

# **Vitamin B12 may inhibit RNA-dependent-RNA polymerase activity of nsp12 of the COVID-19 Virus**

Naveen Narayanan<sup>1,2</sup> and Deepak T. Nair<sup>1</sup>

<sup>1</sup>Laboratory of Genomic Integrity and Evolution, Regional Centre for Biotechnology,  
NCR Biotech Science Cluster, 3<sup>rd</sup> Milestone, Faridabad-Gurgaon Expressway,  
Faridabad- 121001. Haryana.

<sup>2</sup>Manipal Academy of Higher Education, Manipal 576104, Karnataka. India

**KEYWORDS:** nsp12, RNA-dependent-RNA polymerase, COVID-19, inhibitor, Vitamin B12

ABSTRACT: COVID-19 is the causative agent for the ongoing pandemic, and this virus belongs to the Coronaviridae family. Like other members of this family, the virus possesses a positive-sense single-stranded RNA genome. The genome encodes for the nsp12 protein, which houses the RNA-dependent-RNA polymerase (RdRP) activity responsible for the replication of the viral genome. A homology model of nsp12 was prepared using the structure of the SARS nsp12 (6NUR) as a model. The model was used to carry out *in silico* screening to identify molecules among natural products, or FDA approved drugs that can potentially inhibit the activity of nsp12. This exercise showed that vitamin B12 (methylcobalamin) may bind to the active site of the nsp12 protein. A model of the nsp12 in complex with substrate RNA and incoming NTP showed that Vitamin B12 binding site overlaps with that of the incoming nucleotide. A comparison of the calculated energies of binding for RNA plus NTP and methylcobalamin suggested that the vitamin may bind to the active site of nsp12 with significant affinity. It is, therefore, possible that methylcobalamin binding may prevent association with RNA and NTP and thus inhibit the RdRP activity of nsp12. Overall, our computational studies suggest that methylcobalamin form of vitamin B12 may serve as an effective inhibitor of the nsp12 protein.

## Introduction

The members of the Coronaviridae family are viruses with positive-sense single-stranded RNA genomes (1). At present, some members of this family, such as SARS, MERS, and COVID-19 represent pathogens of great concern to public health. An outbreak of COVID-19, which started in the city of Wuhan in China, has now spread to more than 150 countries. At present, there are more than 250000 confirmed cases of the disease caused by this pathogen, with more than 10000 fatalities (2). The WHO has recently classified COVID-19 as a pandemic and asked all countries to do everything necessary to prevent the spread of the disease. At present, there are no effective treatments available against COVID-19, and current medical protocols involve isolating the patient and provide symptomatic treatment for patients with mild disease and oxygen therapy/ventilator support for patients with severe disease.

The genome of COVID-19 is roughly 30 kB long with a gene at the 5' end known as *orf1ab* that encodes for all the polyprotein bearing all the non-structural proteins (3). The virus also possesses genes that code for structural proteins, namely spike (S), envelope (E), membrane (M) and nucleocapsid (N) (4). The polyprotein arising from *orf1ab* may undergo proteolytic processing to give rise to 16 proteins namely nsPs 1-16 (5). Among the cleaved products of the ORF1Ab polyprotein, the proteins of known function include nsp3 which has an adenosine diphosphate-ribose 1"-phosphatase activity (6). The protease activity that is responsible for the cleavage of the polyprotein is present in the nsp5 protein. The nsp12 protein houses the RNA-dependent RNA polymerases that is responsible for duplication of the genome. The RNA helicase activity that is critical for genome duplication is present in the nsp13 protein. Exoribonuclease (exoN) and N7-methyltransferase activities are present in the nsp14 protein (7). The nsp15 protein houses a Nidoviral ribonuclease specific for U, and the nsp16 protein has a SAM-dependent O-methyltransferase activity (3).

There is an urgent need for the identification of new molecules that can reduce viral titers and thus limit the severity of the disease. Towards this end, we have generated a model of the nsp12 molecule from COVID-19 (Co19-nsp12) and used this model to carry out *in silico* screening to identify potential inhibitors of the RNA-

dependent-RNA polymerase activity. Our studies suggest that the methylcobalamin form of Vitamin B12 may serve as an effective inhibitor of the RdRp activity of nsp12.

## **Methods**

### **Homology Modelling of nsp12 and Model of the functional ternary complex**

The sequence corresponding to nsp12 from a sequence of COVID-19 deposited in Genbank by the CDC (Atlanta, USA) with the accession no. MT044257.1 was used. The nsp12 protein from SARS-CoV exhibits 97% identity with the corresponding protein from COVID-19. A homology model was generated using the SWISSMOD server (8), and the structure of nsp12 from the nsp12-nsp7-nsp8 complex (6NUR) of SARS-CoV was used as a template (9). The validation of the model was carried out by the SWISS MOD server (8).

To generate a computational model of the functional ternary complex (Co-19-nsp12:RNA:NTP), initially DALI searches were carried with the model of apo- structure to identify structural orthologues of nsp12 bound to RNA and incoming nucleotide (10). These searches showed that, on superimposition, the Q-beta replicase exhibits significant structural homology in the palm domain. The RNA and incoming nucleotide of the transformed coordinates of the ternary complex of Q-beta replicase (3AVX) were transferred onto the homology model of Co19-nsp12 (11). The local structure in the active site was altered manually to ensure co-ordination of the co-factor ion by the catalytic residues and the triphosphate moiety of the incoming nucleotide as seen previously for DNA polymerases (12). The structure prepared in this way was subjected to energy minimization using DESMOND module of the Schrödinger suite. The structure was minimized in an orthorhombic box containing single point and charge water model and subjected to steepest descent and LBGFS vectors minimization until the difference in energy converged to 0.1 kcal/mol (13).

### ***In Silico* Screening**

The model of Co19-nsp12 was imported into MAESTRO interface of SCHRODINGER suite and prepared using protein preparation wizard program. The prepared structure was used to identify potential binding sites using the SITEMAP

program, which uses site score function to rank the possible binding sites according to size, functionality, and extent of solvent exposure on the protein (14). Sites with a site score value of  $\geq 0.8$  were identified and further examined using PyMOL to see if it could inhibit the RNA binding. Residues spanning the most relevant site were used to prepare a receptor grid using the Receptor Grid Generation module in Schrödinger.

Concurrently various annotated libraries FDA approved drugs, natural products, antiviral compounds and drug repurposing compounds were downloaded from selleckchem.com in the SDF format (15). The downloaded files were converted to accurate energy minimized 3D molecular structures using LIGPREP module in SCHRODINGER. LIGPREP was used to expand tautomeric states, ionization states, ring conformations, and stereoisomers of ligand molecules to produce broad chemical and structural diversity from each molecule along with the correction of Lewis structures and ligand order of these molecules.

Using the glide docking module the prepared grid for the binding site in protein and the library of prepared ligands from LIGPREP were docked. GLIDE generates multiple poses for the ligands, which are initially filtered by spatial fit onto proteins active site and are checked for the complementarity of interaction using ChemScore function. The poses that pass the initial filter are minimized with respect to the receptor grid using OPLS-AA non-bonded ligand-receptor interaction energy. Once the energy is calculated GLIDEScore multi-ligand scoring function assigned scores to the poses. GLIDE docking was carried out in standard-precision (SP) mode, and the molecules that bind to the receptors with good docking score and negative binding energy were used for further analysis.

### **Minimization of the ternary complex of nsp12 and the nsp12-methylcobalamin complex.**

The Co19-nsp12:methylcobalamin complex were then subjected to energy minimization using the DESMOND module of SCHRODINGER suite. The system setup program was used to set up an orthorhombic boundary box containing a simple point-charge water model and 11 Na<sup>+</sup> ions to neutralize the system. This system was then minimized using steepest descent and LBGFS vectors with a convergence threshold of

0.1 kcal/mol. The minimized models of the ternary complex of nsp12 (Co19-nsp12:RNA:GTP:2 Ca<sup>2+</sup> ions) and the nsp12:methylcobalamin complex were analyzed using CONTACT program in CCP4 suite to identify the residues involved in interactions with the substrates and the vitamin, respectively (16). The models of the complexes were superimposed using the COOT program (17) and all figures were prepared using PYMOL (Schrödinger Inc.)

### **Comparison of binding energies**

The binding energy of Co19-nsp12 with substrate or methylcobalamin were calculated by Molecular Mechanics energies combined with Generalized Born and Surface Area continuum solvation (MMGBSA) program in the Schrödinger suite. The docked protein and ligand complex were separated manually and loaded as receptor or ligand, respectively in the MMGBSA module. MMGBSA program minimizes each of them separately as well as in combination using VSGB 2.0 (Variable dielectric Surface Generalized Born) solvation model. The energy of binding is then calculated by subtracting the energy of the optimized free receptor and optimized free ligand from the energy of the optimized complex. Further, to check if methylcobalamin could compete with the natural substrates, the binding energy between the modelled nsp12 and RNA plus incoming nucleotide in the model of the ternary complex was also calculated.

## **Results**

### **Model of the Co19-nsp12 in its apo- and functional state**

The Co19-nsp12 protein shows 97% identity with the corresponding protein from SARS (Figure 1). The structure of the SARS-nsp12 in complex with nsp7 and nsp8 was determined using Cryo-EM and deposited in the PDB with the accession code 6NUR. The structure of nsp12 from 6NUR was used to generate a computational model of the Co19-nsp12 in its apo- state using the SWISSMOD server (Figure 2). The server showed that the stereochemistry of the model was good, with 98% residues in the allowed regions and less than 1% residues in the disallowed regions. Swissmod uses a QMEAN (qualitative model energy analysis) to evaluate the models and the score obtained for the model was -0.65, which is good for further analysis. The model

encompasses residues 4509-5311 of the polyprotein translated from *orf1ab* of the COVID-19 genome and also includes two Zn<sup>2+</sup> ions. The model shows the presence of the N-terminal extension (119-397), fingers (398-581 and 628-687), palm (582-627 and 688-815), and thumb (816-919) domains (Figure 2).

To generate the structure of the Co19-nsp12 structure in its functional state, initially, a DALI search was carried out to identify structural orthologs of this enzyme. The list of enzymes that showed good superimposition with the nsp12 model was analyzed to identify structures of functional ternary complexes. The structure of Q-beta replicase in complex with RNA and incoming nucleotide (3AVX) gave a Z-score of 3.7 and an rmsd of 3.0 in the DALI search. The superimposition was used to generate a model of the Co19-nsp12 protein in complex with RNA bearing template C, incoming nucleotide (dGTP) and two Ca<sup>2+</sup> ions. The model was subjected to energy minimization and converged to a minimum energy of  $-3.97 \times 10^5$  kcal/mol. The Ramachandran plot of the minimized model showed that only 1.75% residues were in the disallowed regions. The model showed the presence of the expected octahedral co-ordination of one of the cofactor ion between the triphosphate moiety and the catalytic residues D762 and D620 (Supplementary Figure 1).

### ***In Silico* Screening identifies Vitamin B12 as a potential binder**

The site map program showed four possible binding sites in the Co19-nsp12 protein, which were ranked according to their ability to bind to various ligands. Site 2 with a site score of 0.99 was selected over site 1 with a site score of 1.03 as it overlapped with the RNA binding groove and catalytic site of the enzyme. The residues spanning site2 were used to generate a receptor grid used for molecular docking. Molecular docking was carried out using annotated libraries of molecules that could bind to Co19-nsp12 open complex using glide dock program. The top hits from each of these libraries were ranked according to their docking score, glide gscore and glide energy based on the interaction between the protein and ligand. Vitamin B12 showed a significant docking score (-8.193), and since it is already part of many drug formulations, the vitamin molecule was selected for further analysis. It was observed that the docked B12 molecule was a Cobalt free version of the vitamin.

Consequently, the methylcobalamin ligand from the complex with human MMACHC (3SC0) was manually docked at the appropriate site in the nsp12 enzyme using the complex model output by GLIDE as a guide (18, 19). The Co19-nsp12:methylcobalamin complex model prepared this way was then subjected to energy minimization. The Co19-nsp12:methylcobalamin converged to a minimum energy of  $-4 \times 10^5$  kcal/mol. The final model of the complex shows the presence of one molecule of methylcobalamin bound in the active site of the nsp12 enzyme (Figure 3).

### **Comparison of interactions in Co19-Nsp12:Vitamin B12 complex and the functional ternary complex of Co19-Nsp12**

The model of the ternary complex of Co19-nsp12 was analyzed to identify the interacting residues. The residues that form Van der Waal interactions with RNA are Lys413, Asn498, Asn499, Lys502, Ser503, Lys513, Lys547, Tyr548, Val559, Arg571, Lys579, Ala582, Arg585, Ile591, Gly592, Thr593, Ser594, Lys595, Phe596, Tyr597, Ser684, Gly685, Asp686, Ala687, Ala690, Tyr691, Leu760, Ser761, Asp762, Asp763, Glu813, Cys815, Ser816, Gln817, Pro834, Asp835, Arg838, Ile839, Val850, Leu856, Glu859, Arg860, Val862, Ser863, Leu864, Ile866, Asp867, Asn913, Arg916, and Tyr917. The residues Asn498, Asn499, Lys502, Ser503, Lys513, Arg571, Lys579, Ala582, Arg585, Thr593, Ser594, Lys595, Tyr597, Cys815, Ser816, Asn913, Arg916, and Tyr917 form polar interactions with RNA. In the ternary complex, the residues Lys574, Asp620, Tyr621, Pro622, Lys623, Asp625, Ser684, Asn693, and Asp762 form polar interactions with the incoming nucleotide i. e. GTP. A624 and G685 form Van der Waal interactions with GTP. The residues Asp620, Tyr621, Asp762, Asp763, and Glu813 also interact with the incoming nucleotide through the co-factor ions. Asp762, Asp763, and Asp620 are the catalytic residues that are responsible for nucleic acid synthesis reaction.

The Co19-nsp12:methylcobalamin complex was analyzed to identify the residues that interact with the vitamin (Figure 4). The nsp12 residues that form Van der Waal contacts with the methylcobalamin molecule include Val168, His441, Tyr457, Tyr460, Ile550, Ser551, Ala552, Lys553, Arg555, Arg557, Thr558, Lys623, Asp625, Arg626, Ser761, Asp762, Glu813, Cys815, Ser816, and Arg838. The residues Lys553, Arg555,



Thr558, Lys623, Asp625, Arg626, Asp762, Glu813, and Arg838 also form polar interactions with the vitamin B12 molecule. One of the three catalytic residues (Asp762) forms polar interactions with the methylcobalamin molecule. A superimposition of the model of the ternary complex and that of the nsp12:methylcobalamin complex was carried out to ascertain the level of overlap between the binding sites of the natural substrates of nsp12 and the methylcobalamin molecule. The superimposition showed that the binding site of methylcobalamin overlaps with that of the incoming nucleotide and the terminal primer nucleotide of the RNA substrate (Figure 5).

### **Energy of binding of Co19-nsp12 with natural substrates versus methylcobalamin**

Using the MMGBSA program, the energy of interaction between Co19-nsp12 and RNA+GTP was compared with that of between nsp12 and methylcobalamin. The calculated binding energy for the interaction of RNA and incoming nucleotide with Co19-Nsp12 is -95 kcal/mol. In comparison, the calculated binding energy value for the interaction of methylcobalamin with Co19-nsp12 is -48 kcal/mol. These values suggest that Vitamin B12 can bind with significant affinity at the active site of the Co19-nsp12 enzyme. Therefore, the binding of methylcobalamin may inhibit the formation of functional ternary complex of nsp12 and thus prevent replication of the viral genome.

### **Discussion**

The Co19-nsp12 shows 97% sequence identity with the corresponding protein from SARS. Due to the availability of the structure of the complex of nsp12-nsp7-nsp8 structure from SARS, a working model of the Co19-nsp12 could be generated. The model of the apo-structure was used to generate a structure of the functional ternary complex of the Co19-nsp12 protein. *In silico* screening, using the model of Co19-nsp12 suggests that methylcobalamin may bind to the active site of the enzyme. The vitamin binds in the region where the incoming nucleotide binds with high affinity. Also, the binding site of the vitamin overlaps with that of the incoming nucleotide and substrate RNA.

Consequently, the binding of methylcobalamin may prevent the binding of incoming nucleotide and substrate RNA and incoming nucleotide due to steric clashes. The

studies presented, therefore, suggest that methylcobalamin may be a possible inhibitor of the RNA-dependent-RNA polymerase activity of the Co19-nsp12 enzyme. Since this enzyme is critical for the replication of the viral enzyme, the inhibition of this enzyme can result in lower viral titres and reduced disease morbidity. Because the number of infected patients are increasing daily and that about 7% of them are in serious condition, the ability of methylcobalamin to inhibit viral replication should be tested urgently using *in vitro* and *in vivo* assays. In addition, given the urgency of the situation and the fact that methylcobalamin is already part of drug formulations, doctors may consider adding or increasing the dosage of methylcobalamin in their current patient care protocols.

## Reference

- [1] Modrow, S., Falke, D., Truyen, U., Schätzl, H., Modrow, S., et al. (2013) Viruses with Single-Stranded, Positive-Sense RNA Genomes. In *Molecular Virology*. . pp. 185–349, Springer Berlin Heidelberg.
- [2] Johns Hopkins Coronavirus Resource Center.
- [3] Wu, F., Zhao, S., Yu, B., Chen, Y.-M., Wang, W., et al. (2020) A new coronavirus associated with human respiratory disease in China. *Nature*.
- [4] Shang, W., Yang, Y., Rao, Y., and Rao, X. (2020) The outbreak of SARS-CoV-2 pneumonia calls for viral vaccines. *npj Vaccines* 5, 18.
- [5] Graham, R. L., Sparks, J. S., Eckerle, L. D., Sims, A. C., and Denison, M. R. (2008) SARS coronavirus replicase proteins in pathogenesis. *Virus Research* 133, 88–100.
- [6] Saikatendu, K. S., Joseph, J. S., Subramanian, V., Clayton, T., Griffith, M., et al. (2005) Structural basis of severe acute respiratory syndrome coronavirus ADP-ribose-1"-phosphate dephosphorylation by a conserved domain of nsP3. *Structure* 13, 1665–1675.
- [7] Ma, Y., Wu, L., Shaw, N., Gao, Y., Wang, J., et al. (2015) Structural basis and functional analysis of the SARS coronavirus nsp14-nsp10 complex. *Proceedings of the National Academy of Sciences of the United States of America* 112, 9436–9441.

- [8] Biasini, M., Bienert, S., Waterhouse, A., Arnold, K., Studer, G., et al. (2014) SWISS-MODEL: modelling protein tertiary and quaternary structure using evolutionary information. *Nucleic Acids Research* 42, W252–W258.
- [9] Kirchdoerfer, R. N., and Ward, A. B. (2019) Structure of the SARS-CoV nsp12 polymerase bound to nsp7 and nsp8 co-factors. *Nat Commun* 10, 2342–2342.
- [10] Holm, L., and Laakso, L. M. (2016) Dali server update. *Nucleic Acids Res* 44, W351--5.
- [11] Takeshita, D., and Tomita, K. (2012) Molecular basis for RNA polymerization by Q beta replicase. *Nat.Struct.Mol.Biol.* 19, 229–237.
- [12] Kottur, J., and Nair, D. T. Pyrophosphate hydrolysis is an intrinsic and critical step of the DNA synthesis reaction. *Nucleic Acids Research*.
- [13] Bowers, K. J., Chow, E., Xu, H., Dror, R. O., Eastwood, M. P., et al. (2006) Scalable algorithms for molecular dynamics simulations on commodity clusters. In *Proceedings of the 2006 ACM/IEEE Conference on Supercomputing, SC'06*. . p. 84, ACM Press, New York, New York, USA.
- [14] Halgren, T. (2007) New method for fast and accurate binding-site identification and analysis. *Chemical Biology and Drug Design* 69, 146–148.
- [15] Selleckchem.com - Inhibitor Expert (Inhibitors, Compound Libraries).
- [16] Winn, M. D., Ballard, C. C., Cowtan, K. D., Dodson, E. J., Emsley, P., et al. (2011) Overview of the CCP4 suite and current developments. *Acta Crystallogr D Biol Crystallogr* 67, 235–242.
- [17] Emsley, P., and Cowtan, K. (2004) Coot: model-building tools for molecular graphics. *Acta Crystallogr D Biol Crystallogr* 60, 2126–2132.
- [18] Koutmos, M., Gherasim, C., Smith, J. L., and Banerjee, R. (2011) Structural basis of multifunctionality in a vitamin B12-processing enzyme. *J.Biol.Chem.* 286, 29780–29787.
- [19] Friesner, R. A., Banks, J. L., Murphy, R. B., Halgren, T. A., Klicic, J. J., et al. (2004) Glide: A New Approach for Rapid, Accurate Docking and Scoring. 1. Method and Assessment of Docking Accuracy. *Journal of Medicinal Chemistry* 47, 1739–1749.

## Author contribution

DTN and NN conceived the project. NN carried out the computations and DTN supervised the work. DTN and NN analyzed the results and wrote the manuscript.

**Competing Interests statement:** None declared

## Figure Legends:

**Figure 1:** Alignment of the sequence of Covid19-nsp12 (119-901) with the sequence of the available structure of SARS-nsp12 (6NUR). The Covid19-nsp12 sequence exhibits about 97% identity with the corresponding stretch in SARS-nsp12.

**Figure 2:** Model of the Covid19-nsp12 enzyme in its apo- state. The N-terminal extension region and the palm, fingers and thumb domains are shown in green, cyan, yellow and orange, respectively. The catalytic residues Asp620, Asp762 and Asp763 are coloured according to element and shown in stick representation.

**Figure 3:** Model of the Covid19-nsp12 enzyme in complex with methylcobalamin. The Vitamin B12 molecule is shown in stick representation and coloured according to element. The N-terminal extension region and the palm, fingers and thumb domains are shown in green, cyan, yellow and orange, respectively. The catalytic residues Asp620, Asp762 and Asp763 are coloured according to element and shown in stick representation.

**Figure 4:** Residues of nsp12 that interact with methylcobalamin. The Vitamin B12 molecule is shown in stick representation and coloured according to element. The N-terminal extension region and the palm, fingers and thumb domains are shown in green, cyan, yellow and orange, respectively. The interacting residues are coloured according to element and shown in stick representation.

**Figure 5:** Binding site of methylcobalamin overlaps with that of incoming nucleotide. (A) Superimposition of the models of the functional ternary complex (cyan) and that of nsp12:methylcobalamin (green) is displayed. The protein chains are shown in ribbon representation. The catalytic residues, incoming GTP and methylcobalamin are shown in stick representation and the cofactor ions are shown as spheres (B) The surface of the protein molecule is displayed and the RNA, incoming nucleotide, and methylcobalamin are shown in stick representation and coloured white, red and

magenta, respectively. The vitamin B12 binding site overlaps with that of the incoming nucleotide and the terminal primer nucleotide of the RNA substrate.

**Supplementary Figure Legends:**

**Supplementary Figure 1:** Model of the functional ternary complex of the Co19-nsp12 enzyme in complex. The RNA substrate and incoming GTP are shown in stick representation and coloured according to element. The N-terminal extension region and the palm, fingers and thumb domains are shown in green, cyan, yellow and orange, respectively. The catalytic residues Asp620, Asp762 and Asp763 are coloured according to element and shown in stick representation and the cofactor ions are shown as blue spheres.

Covid19_nsp12 SARS_6NUR	119	178
	QRLTKYTMADLVYALRHFDEGNCDTLKEILVITYNCCDDYFNKKDWYDFVENPDI LRVYA	
	QRLTKYTMADLVYALRHFDEGNCDTLKEILVITYNCCDDYFNKKDWYDFVENPDI LRVYA	
Covid19_nsp12 SARS_6NUR	179	218
	NLGERVRQALLKTVQFCDAMRNAGIVGVLTLDNQDLNGNWYDFGDFIQTTPGSGVPIVDS	
	NLGERVRQSLLKTVQFCDAMRDAGIVGVLTLDNQDLNGNWYDFGDFVQVAPGCGVPIVDS	
Covid19_nsp12 SARS_6NUR	219	278
	YYSLLMPILTLTRALTAESHVDTDLTKPYIKWDLKLYDFTEERLCLFDRIYFKYWDQTYHP	
	YYSLLMPILTLTRALAAESHMDADLAKPLIKWDLKLYDFTEERLCLFDRIYFKYWDQTYHP	
Covid19_nsp12 SARS_6NUR	279	338
	NCVNCILDDRCILHCANFNVLFSVFPPTSEGPLVRKIFVDGVPFVVS TGYHFRELGVVHN	
	NCINCLDDRCILHCANFNVLFSVFPPTSEGPLVRKIFVDGVPFVVS TGYHFRELGVVHN	
Covid19_nsp12 SARS_6NUR	339	398
	QDVNLHS SRLSFKELLVYAADPAMHAASGNLLDKRTTCSVAAL TNNVAFQTVKPGNFN	
	QDVNLHS SRLSFKELLVYAADPAMHAASGNLLDKRTTCSVAAL TNNVAFQTVKPGNFN	
Covid19_nsp12 SARS_6NUR	399	458
	KDFYDFAVSKGFFKEGSVELKHFFFAODGNAAISDYDYRYNLP TMCDIRQLLFVVEVV	
	KDFYDFAVSKGFFKEGSVELKHFFFAODGNAAISDYDYRYNLP TMCDIRQLLFVVEVV	
Covid19_nsp12 SARS_6NUR	459	518
	DKYFDCYDGGCINANQVIVNNLDKSAGFPFNKVGKARLYYD SMSYEDQDALFAYTKRNV I	
	DKYFDCYDGGCINANQVIVNNLDKSAGFPFNKVGKARLYYD SMSYEDQDALFAYTKRNV I	
Covid19_nsp12 SARS_6NUR	519	578
	PTITQMNLKYAISAKNRARTVAGVSI CSTMTRQFHQKLLKSI AATR GATVVI GTSKFYG	
	PTITQMNLKYAISAKNRARTVAGVSI CSTMTRQFHQKLLKSI AATR GATVVI GTSKFYG	
Covid19_nsp12 SARS_6NUR	579	638
	GWHNMLKTVYSDVENPHLMGWDYPKADRAMPNMLRIMASLVLARKHNTCCSLSHRFYRLA	
	GWHNMLKTVYSDVETPHLMGWDYPKADRAMPNMLRIMASLVLARKHNTCCSLSHRFYRLA	
Covid19_nsp12 SARS_6NUR	639	698
	NECAQVLSEMVMCGGSLYVKPGGTS SGDAT TAYAN SVFNICQAVTANVNALLS TDGNKIA	
	NECAQVLSEMVMCGGSLYVKPGGTS SGDAT TAYAN SVFNICQAVTANVNALLS TDGNKIA	
Covid19_nsp12 SARS_6NUR	699	758
	DKYVRNLQHRLYECLYRNRDVD TDFVNEFYAYLRKHF SMMI LSDDAVVC FNSTYASQGLV	
	DKYVRNLQHRLYECLYRNRDVDHEFVDEFYAYLRKHF SMMI LSDDAVVC VNSNYAAQGLV	
Covid19_nsp12 SARS_6NUR	759	818
	ASIKNFKSVLYYQNNVFMSEAKCWTE TDLTKGPHEFC SQHTMLVKQGDDYVYLPYDPSPR	
	ASIKNFKAVLYYQNNVFMSEAKCWTE TDLTKGPHEFC SQHTMLVKQGDDYVYLPYDPSPR	
Covid19_nsp12 SARS_6NUR	819	878
	IILGAGCFVDDIVKTDGTLMIERFVSLAIDAYPLTKHPNQEYADV FHLYLQYIRKLHDEL T	
	IILGAGCFVDDIVKTDGTLMIERFVSLAIDAYPLTKHPNQEYADV FHLYLQYIRKLHDEL T	
Covid19_nsp12 SARS_6NUR	879	901
	GHMLDMYSVMLTNDNTSRYWEPE	
	GHMLDMYSVMLTNDNTSRYWEPE	

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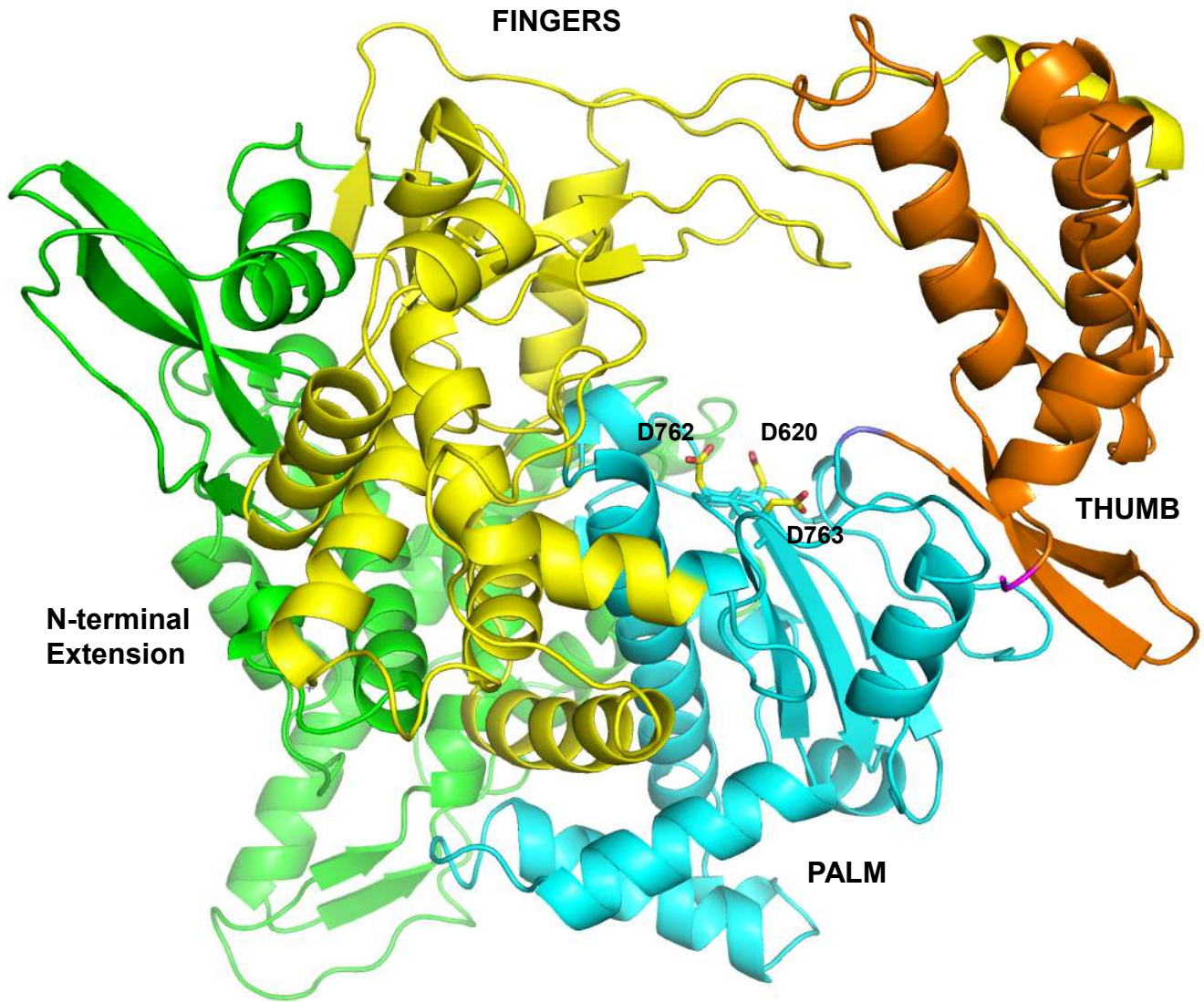


Figure 2: Model of the Covid19-nsp12 enzyme in its apo- state. The N-terminal extension region and the palm, fingers and thumb domains are shown in green, cyan, yellow and orange, respectively. The catalytic residues Asp620, Asp762 and Asp763 are coloured according to element and shown in stick representation.

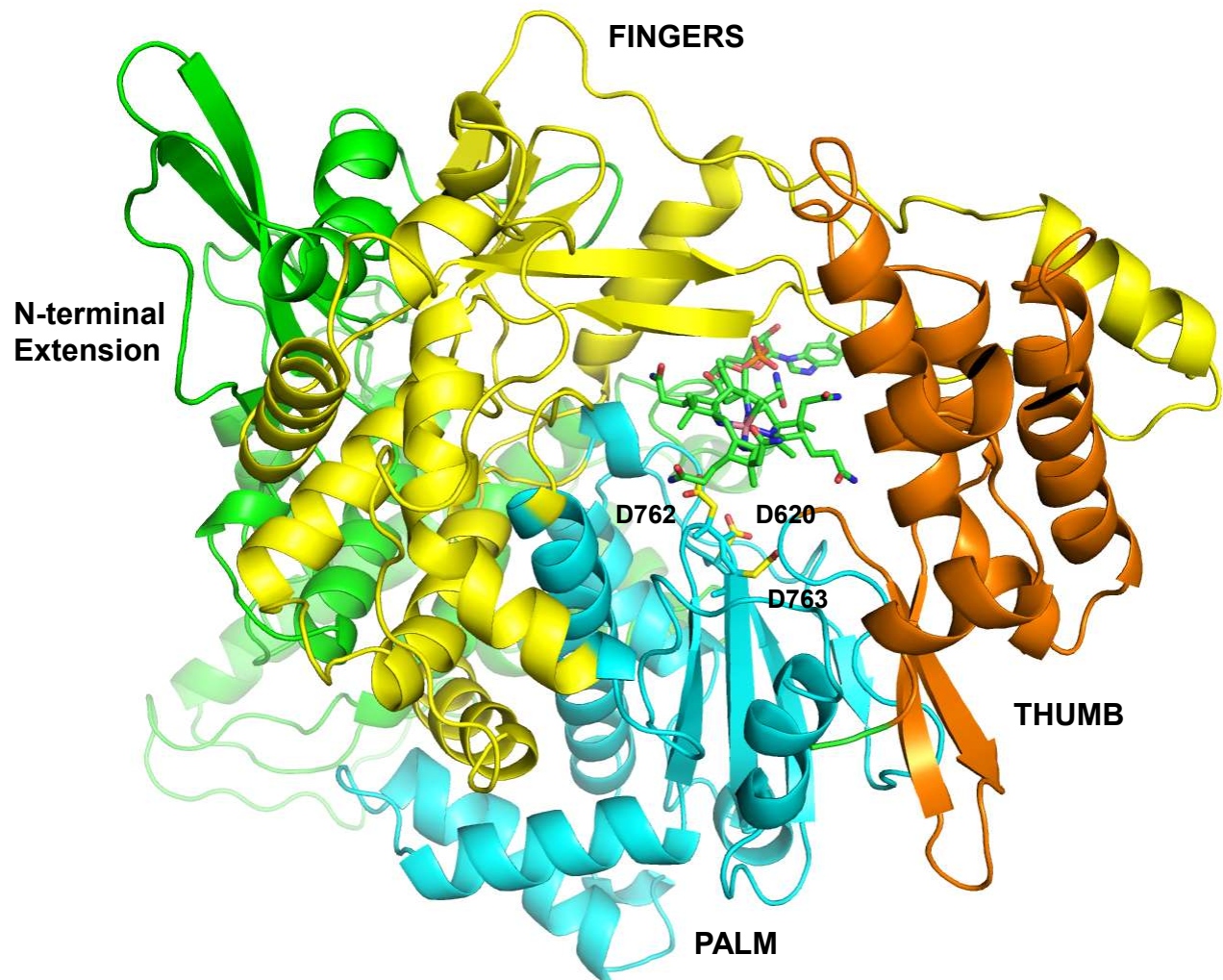


Figure 3: Model of the Covid19-nsp12 enzyme in complex with methylcobalamin. The Vitamin B12 molecule is shown in stick representation and coloured according to element. The N-terminal extension region and the palm, fingers and thumb domains are shown in green, cyan, yellow and orange, respectively. The catalytic residues Asp620, Asp762 and Asp763 are coloured according to element and shown in stick representation.



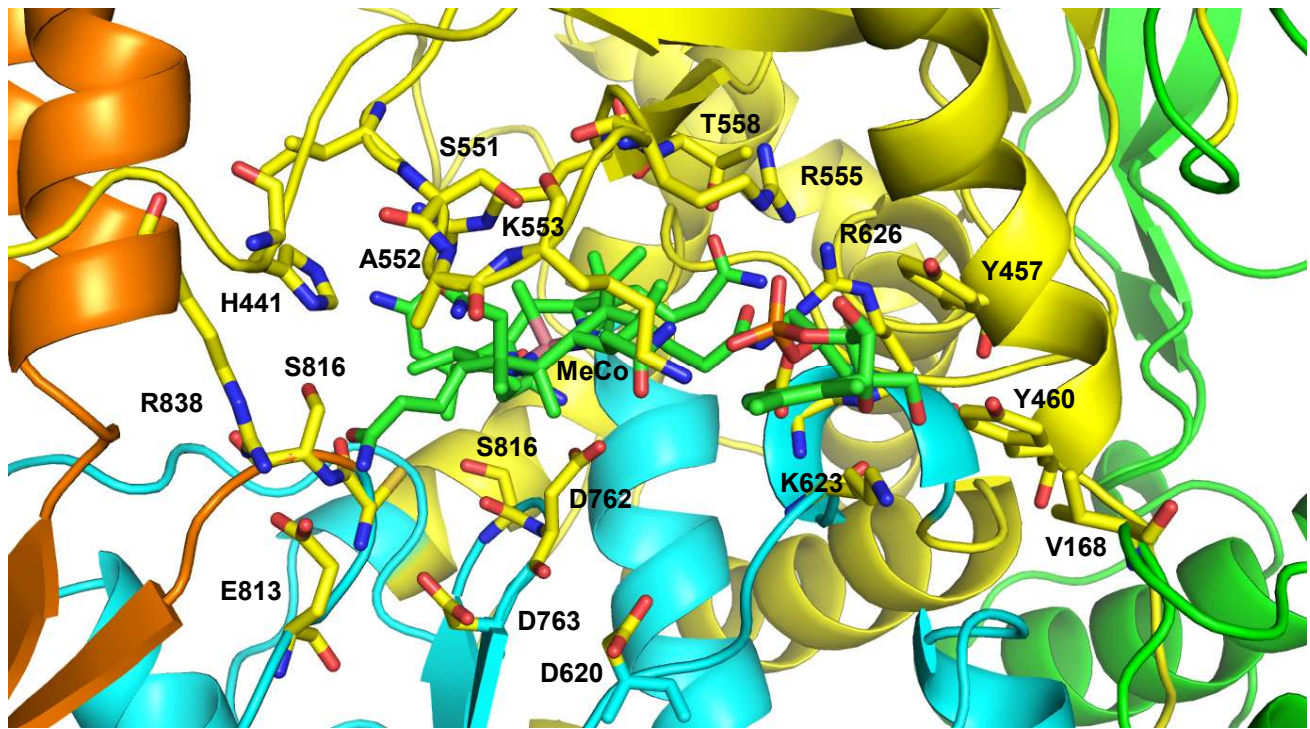


Figure 4: Residues of nsp12 that interact with methylcobalamin. The Vitamin B12 (MeCo) molecule is shown in stick representation and coloured according to element. The N-terminal extension region and the palm, fingers and thumb domains are shown in green, cyan, yellow and orange, respectively. The interacting residues are coloured according to element and shown in stick representation.

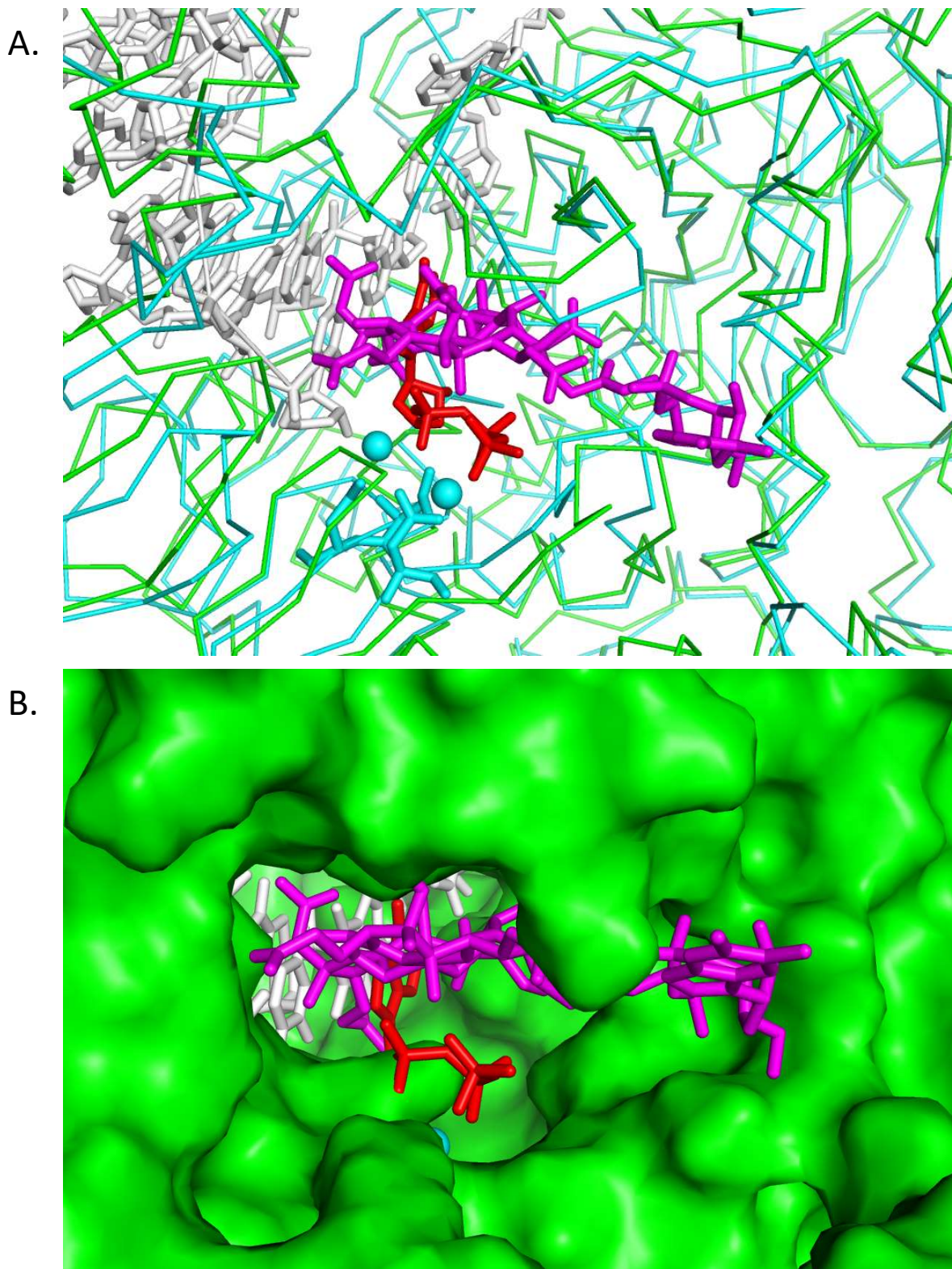
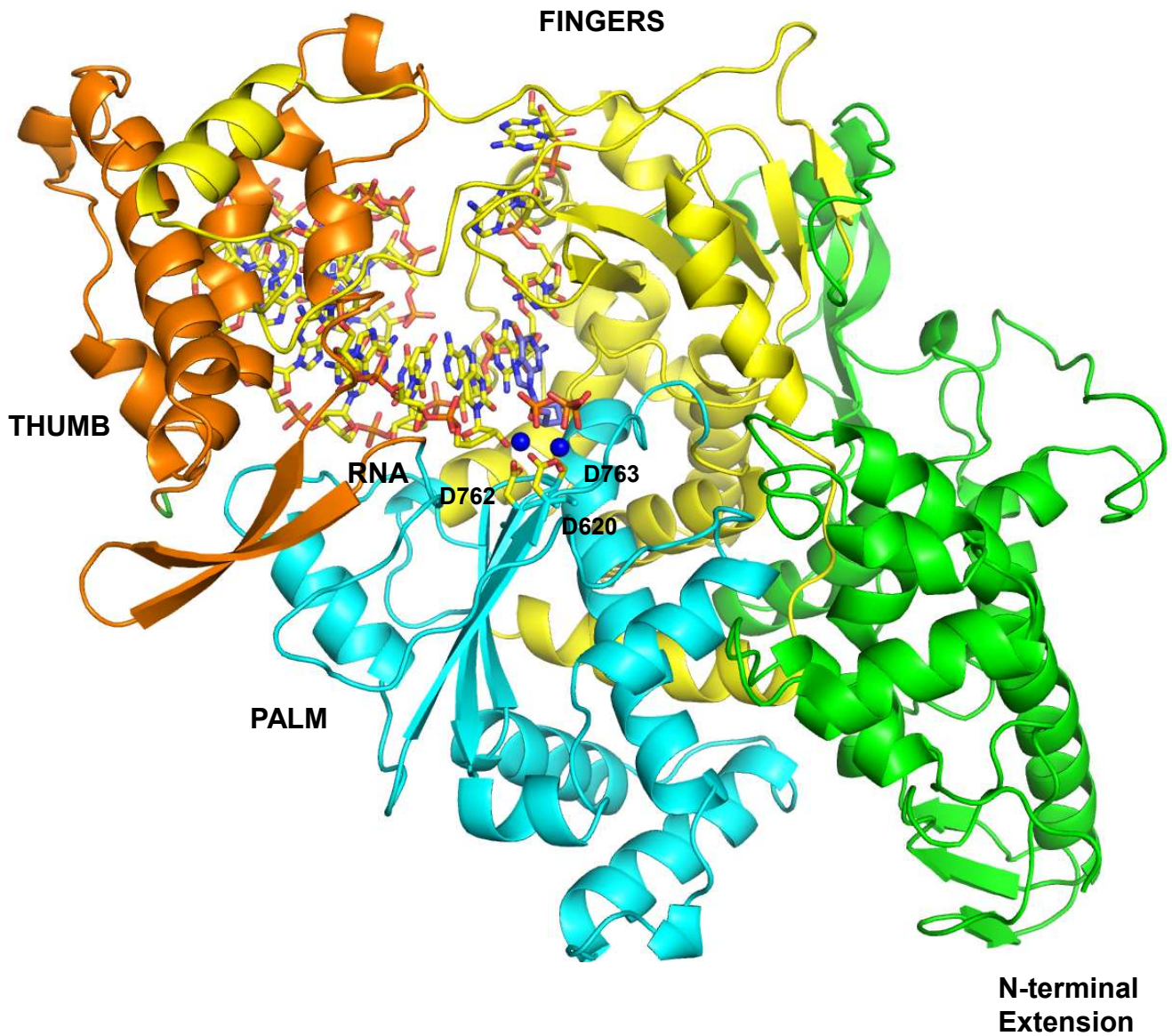


Figure 4: Binding site of methylcobalamin overlaps with that of incoming nucleotide. (A) Superimposition of the models of the functional ternary complex (cyan) and that of nsp12:methylcobalamin (green) is displayed. The protein chains are shown in ribbon representation. The catalytic residues (cyan), incoming GTP (red) and methylcobalamin (magenta) are shown in stick representation and the cofactor ions are shown as spheres (B) The surface of the protein molecule is displayed and the RNA, incoming nucleotide, and methylcobalamin are shown in stick representation and coloured white, red and magenta, respectively. The vitamin B12 binding site overlaps with that of the incoming nucleotide and the terminal primer nucleotide.



Supplementary Figure 1: Model of the functional ternary complex of the Covid19-nsp12 enzyme in complex. The RNA substrate and incoming GTP are shown in stick representation and coloured according to element. The N-terminal extension region and the palm, fingers and thumb domains are shown in green, cyan, yellow and orange, respectively. The catalytic residues Asp620, Asp762 and Asp763 are coloured according to element and shown in stick representation and the cofactor ions are shown as blue spheres.